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14. ABSTRACT Post traumatic stress disorder (PTSD) is a complex clinical disorder resulting from exposure to intense, life-threatening events resulting in persistent re-experiencing of the trauma, avoidance of stimuli associated with the trauma, dissociation, and heightened arousal which severely impact social and occupational functioning. Recent work has underscored morphological and functional brain alterations in PTSD patients using brain imaging with MRI, SPECT and PET imaging. Despite this encouraging preliminary work, there exists only a limited understanding of the pathophysiological changes which may subserve symptoms of PTSD. Preclinical studies now suggest that inflammatory changes may be implicated in neuronal loss in models of PTSD. Microglia represent a key inflammatory cell mediator within the CNS. Upon activation, these cells densely express an 18 kDa translocator protein (TSPO) receptors on their cell surface. Hence, it is possible to develop a radiotracer which targets TSPO as a marker for neuroinflammation. We have performed preliminary work with the TSPO imaging agent 123-I CLINDE with a goal of this proposal is to establish and validate an imaging biomarker for neuroinflammation in PTSD subjects that could both enhance our understanding of the pathophysiology and identify new therapeutic strategies for PTSD.					
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Introduction

Activated microglia have been proposed to play a major role in the pathogenesis of several central nervous system conditions including post-traumatic stress disorder (PTSD). Microglia are a cell type that represents over 10% of the cells in the brain and become “activated” in response to various stimuli. This activation is thought to control areas of inflammation in the brain. In certain neurological diseases, this activation is thought to contribute to progression of neuronal damage. When microglia become activated they express peripheral benzodiazepine receptors (PBR) or binding sites on their mitochondrial membrane. PBRs are functionally and structurally distinct from central benzodiazepine receptors associated with gamma-aminobutyric acid (GABA)-regulated chloride channels. PBRs are found in abundance in peripheral organs and hematologic cells, but are present at only very low levels in the normal central nervous system (Banati, 2002).

A number of ligands with high in vitro affinity to PBR have been investigated for PET or SPECT imaging. To date, 11-C PK-11195 has been considered the best agent, in spite of its limitations (Kropholler, 2007; Turkheimer, 2007; Schuitemaker, 2007; Imaizumi, 2007). 123-I PK-11195 was also evaluated in humans but its low brain penetration makes it unfavorable for further evaluation (Shinotoh, 2006; Dumont, 1999; Gildersleeve, 1996). More recently, 18-F and 11-C labeled DAA1106 analogs were shown to be promising for imaging in nonhuman primates, but to date no report has been published concerning their use in humans (Boutin [65]). PET imaging using 11-C PK11195, a PBR agent, has demonstrated increased uptake in patients with several neurodegenerative conditions. Although 11-C PK11195 has provided critical proof of concept that in vivo imaging may be used to monitor neuroinflammation, there are several limitations to this ligand. Quantitation of 11-C PK11195 is difficult and use of 11-C labeled ligands are not practical in large clinical studies because of the short half-life (20 min).

In this proposal we planned to develop a PBR-binding radiotracer labeled with 123-I (T_{1/2} 13.1 h) to enhance the clinical utility of imaging of neuroinflammatory targets. Unfortunately, 123-I CLINDE, the radiotracer initially proposed had limited signal:noise properties in other human trials. We have subsequently identified a better agent for interrogating PBR in PTSD subjects, 18-F PBR111, a PET compound with more favorable human preliminary data than 123-I CLINDE. PBR111 appears to bind selectively to the PBR, also referred to as TSPO (the 18kDa translocator protein). An increase PBR111 binding to PBR is a potential marker of microglial activation in the CNS. The increase in PBR111 binding may be an indicator of the transition of microglia from a resting to an activated state, reflecting inflammation and its clinical sequelae relevant to PTSD symptomatology.

Body

This is a Phase 1, single-center, open-label, non-randomized, clinical study in PTSD and HCs to evaluate the kinetics, clearance and cerebral distribution of [18F]PBR-111 as outlined in the study protocol on file with the DOD. Investigational new drug approval has been granted by the FDA to study this agent (IND # 107,622). The

underlying goal of this study is to assess [18F]PBR-111 PET imaging as a tool to detect microglial activation in the brain of PTSD research participants and similarly aged healthy subjects. All study procedures will be conducted at the Institute for Neurodegenerative Disorders (IND) in New Haven, CT.

Approximately 8 subjects with post-traumatic stress disorder (PTSD) and 6 similarly aged healthy controls will be recruited to participate in this study. Previous studies with novel radiotracers in healthy and disease cohorts suggests there sample sizes are sufficient to detect 20% differences for outcome measures with good test-retest reproducibility. Healthy controls will be screened to ensure that there is no evidence of significant neurological or psychiatric disturbance.

The study population will consist of male or female subjects of any ethnic group with the features of PTSD and HC subjects. The study investigator will confirm (or reject) the diagnosis of PTSD based on the DSM-IV criteria after clinical and neuro-psychiatric examination. Only subjects fulfilling all inclusion criteria and none of the exclusion criteria for PTSD and healthy controls will be included into the study. To determine the efficacy of [18F]PBR-111 PET scans in subjects with PTSD and HCs on the basis of neocortical tracer binding pattern, the PET scans will be assessed by a nuclear physician experienced in the field of neuro-imaging. PET scan findings will be classified either as abnormal (i.e., significant neocortical uptake in predefined regions) or as normal (i.e. no significant neocortical uptake in predefined regions). The nuclear physician will be unaware of the clinical diagnosis.

In support of this study, we have assessed [18F]-PBR111 as a potential marker for microglial activation in association with neuronal damage in previous human and non-human primate PET studies to determine the suitability of the radiotracer for interrogating multiple central nervous system conditions including PTSD. These studies are briefly described below.

Key Research Accomplishments

We decided to consider additional TSPO agents with better properties prior to exposing PTSD subjects and controls to imaging studies with a poor tracer. In this vein, outside the context and funding of the present proposal, we initiated human studies with three additional TSPO radioligands with the aim of quickly deciding upon a more optimal radioligand for the PTSD work. This report summarizes the progress of this work, even though we realize it is not being supported or part of the research initiatives funded under this grant. In addition, we have contacted the grant administrator to indicate we would wish to request a no cost extension of the work to facilitate the best scientific study possible under the current award. Summarizing this, objective progress in support of the initiation of the PTSD trial includes:

1. We have established collaborations with colleagues in the Yale Department of Psychiatry (Drs. Steven Southwick and John Krystal) for assistance in the referral and evaluation of PTSD research participants.
2. Obtained an IND for 18F-PBR111 for human use.

3. Finalized technical aspects of radiochemistry, PET acquisition, radiometabolite analysis, and image processing and signal quantification in support of the current study.
4. Completed all DOD human research approval requirements.

The issues and work delineated in point 3 above are described in the subsequent portion of this report.

Reportable Outcomes

Review of additional clinical imaging data from studies in controls and selected neuropsychiatric disorders

The following reports preliminary work done with 18F PBR111, unfunded by, but in support of the current study.

A. [18]-F PBR111 PET Baboon studies

PET investigations were conducted involving the bolus injection of [18]-F PBR111 in three ovariectomized female baboons (*Papio anubis*). Briefly, [18]-F PBR111 was prepared by halogen exchange of the appropriate precursor with [18F]fluoride in the presence of potassium carbonate and kryptofix-222 followed by HPLC isolation. Images were acquired after administration of 4.60 to 5.85 mCi of [18]-F PBR111 injected as an intravenous bolus in three female ovariectomized baboons (weight 12.10 – 18.65 kg). The animals were studied under isoflurane anesthesia after ketamine and rubinol induction. Serial dynamic brain PET acquisitions were acquired for up to 3-h post injection. PET studies were analyzed to evaluate the maximal uptake of radioactivity and washout, pattern of metabolite formation in blood, and effect of displacing agent PK11195 on specific binding in different brain regions.

Safety data

Following the induction of anesthesia through the end of the study heart rate, respiratory rate, blood pressure, and temperature were monitored every fifteen minutes. Data from the anesthesia record were entered into a database and to assess alterations in vital signs following injection of [18]-F PBR111. The animal tolerated the injection of [18]-F PBR111 and scan procedures without significant adverse effects. There were no systematic effects noted on vital signs following [18]-F PBR111 injection. Vital sign data from a representative study is indicated in Fig A1 below.

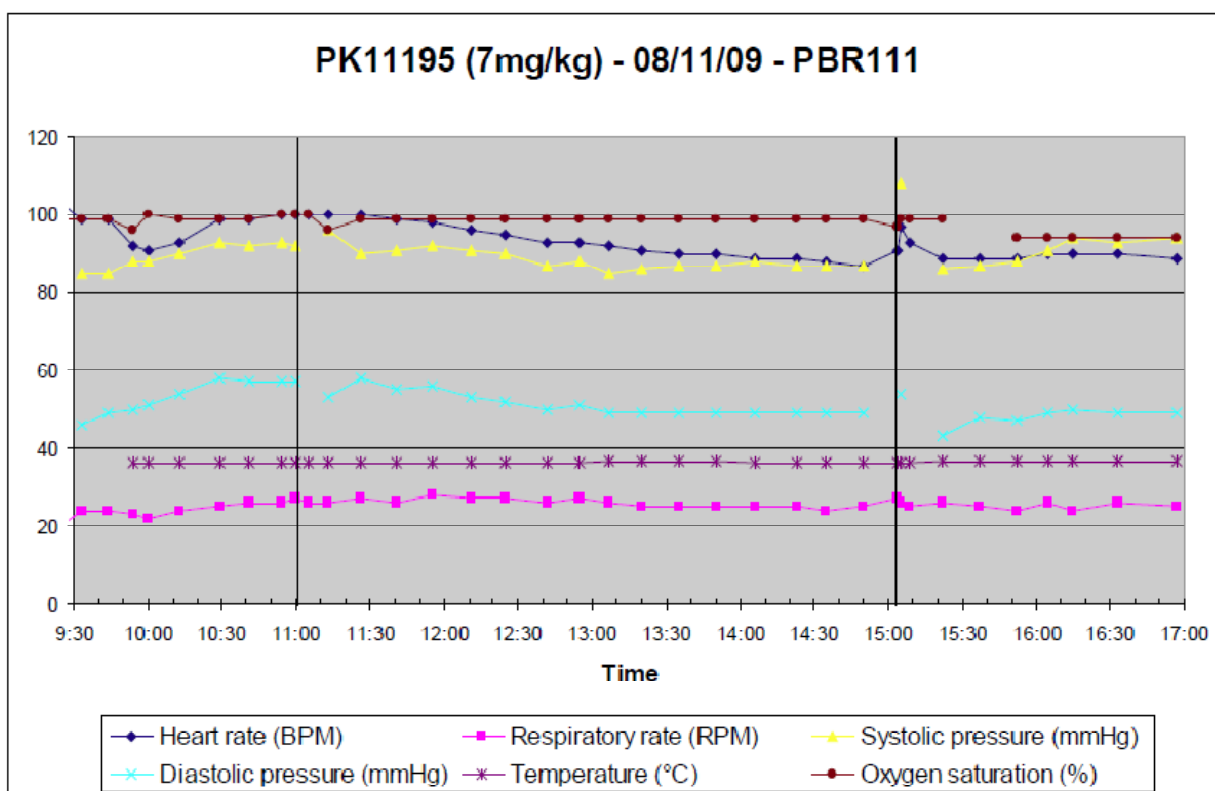


Figure A1. Panel indicates vital signs in a female baboon following intravenous injection of $[^{18}\text{F}]$ -PBR111 (5.04 mCi) and PK11195 (7 mg/kg). The two vertical lines on the graph indicate the time points at which the radiotracer and displacing agent were injected, respectively.

B. $[^{18}\text{F}]$ -PBR111 PET Human studies

A total of 11 human subjects (8 healthy volunteers, 3 Alzheimer's subjects) have been studied with $[^{18}\text{F}]$ -F PBR111 in the context of separately funded and approved investigations to validate and optimize the imaging outcome measure for the current proposal. These studies are performed under IND # 107,622 at the Institute for Neurodegenerative Disorders, New Haven, CT.

Subject ID	Mind #	Gender	Diagnosis	Dose mCi	MBq
PBR111-02	292-07378	M	HC	4.72	174.64
PBR111-01	292-07412	M	HC	4.41	163.17
PBR111-03	292-07428	F	HC	3.95	146.15
PBR111-05	292-07506	M	HC	5.00	185.00
PBR111-08	292-07607	M	HC	3.11	115.07
PBR111-09	292-07621	F	HC	4.97	183.89
PBR111-11	292-07622	F	HC	3.55	131.35
PBR111-12	292-07635	F	HC	5.01	185.37
PBR111-08	292-07658	M	HC	5.01	185.37
PBR111-09	292-07668	F	HC	3.00	111.00
PBR111-05	292-07506	M	HC	4.69	173.53

Subject ID	Mind #	Gender	Diagnosis	Dose	MBq
PBR111-04	292-07474	M	AD	4.59	169.83
PBR111-06	292-07533	M	AD	3.49	129.13
PBR111-13	292-07683	M	AD	4.92	182.04

Table A1. Human PBR111 PET subjects studied to date include 8 healthy volunteers (HC) and 3 Alzheimer's (AD) subjects.

Following bolus injection of [18]-F PBR111, serial dynamic PET brain acquisitions were obtained for three hours. Arterial and venous sampling was obtained for determining the metabolite corrected brain input function. Images were reconstructed with a standard iterative algorithm with scatter, randoms, and attenuation correction applied. Each serial image volume was spatially normalized and co-registered with a T1 weighted MRI and a standardized volume of interest brain template based on the AAL template was checked for accuracy on the MRI then transferred to the PET image for extraction of standard uptake values (SUV). These regional brain SUVs were then modeled with classic two tissue (2T), one tissue (1T), and Logan plot kinetic analyses to determine V_t . In addition, a pixelwise analysis (2T) was performed on the image volumes using modeled constraints determined in the VOI-level analysis.

Findings: All subjects tolerated the PET procedures without subjective or objective changes following radiotracer injection. Specifically, there were no changes in vital signs, EKG measures, serum chemistries, hematological indices, or somatic complaints relative to baseline assessments.

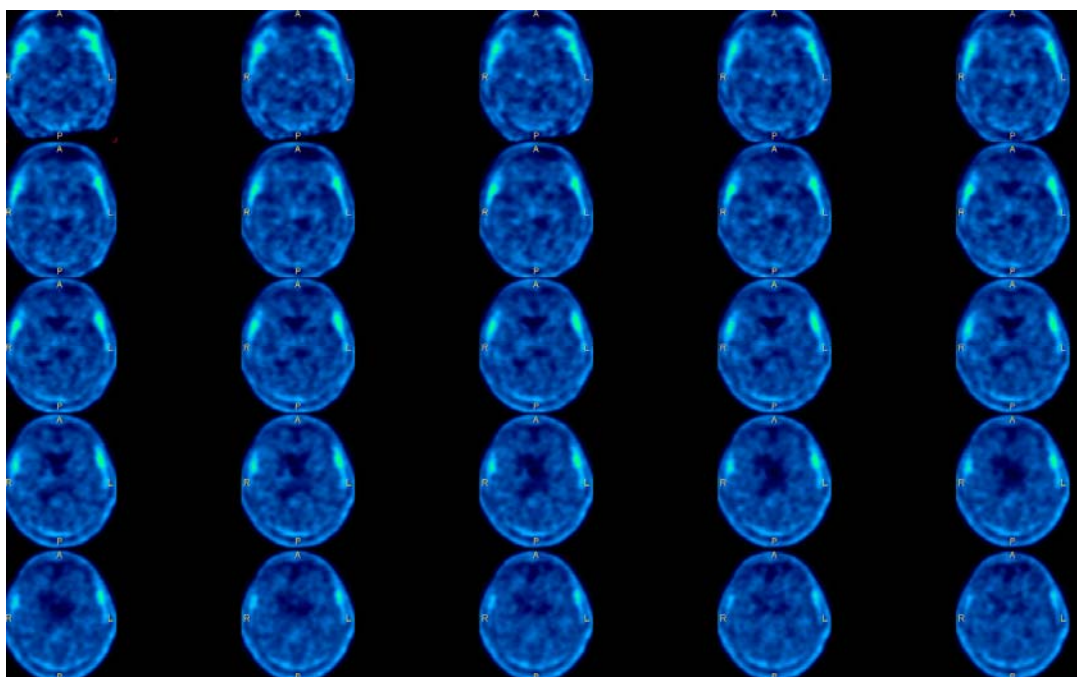


Figure A2: Representative PET axial images obtained from a healthy human volunteer receiving a bolus injection of [18]F PBR111.

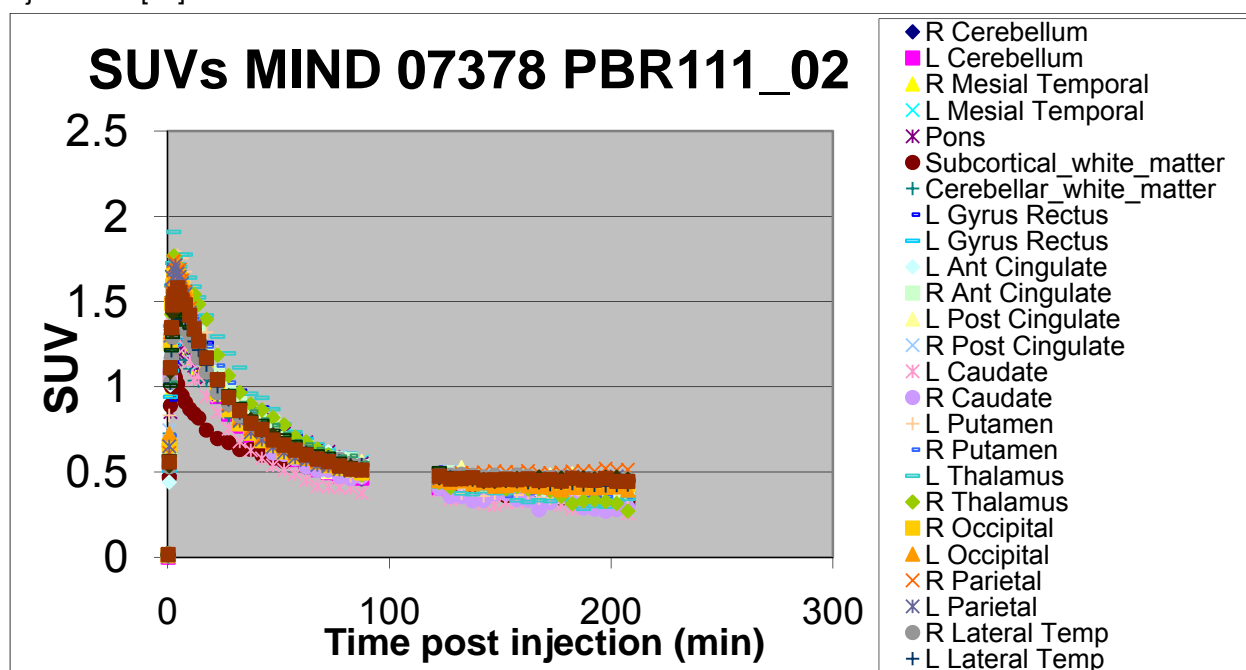


Figure A3: Standard uptake value (SUV) time-activity curve for multiple brain regions in a healthy human subject receiving a bolus injection of [18]F PBR111. The gap in the curve indicates a rest period outside the camera when no images were acquired.

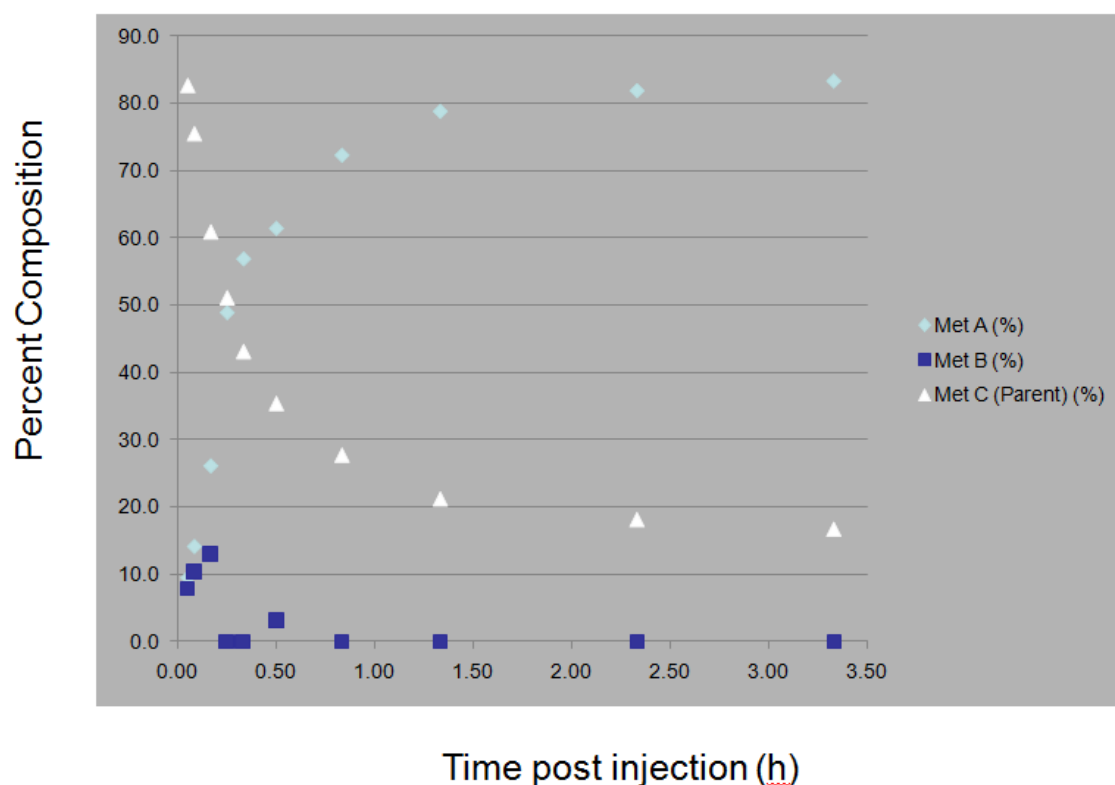


Fig A4: Plasma analysis in healthy human volunteer indicating metabolism of [18]F PBR111 into two metabolites with about 22% of parent compound present at one h post injection.

Analysis of blood data demonstrated the following:

- 1) Venous plasma analysis demonstrates parent compound is quickly metabolized with residual parent reflecting 10-20% of activity at 90 minutes and some variability between subjects.
- 2) PBR111 is primarily metabolized to two extractable metabolites- one of which is predominant (Metabolite A= 50-90%, Metabolite B = 0-30%). No evidence of confounding lipophilic metabolite.
- 3) There are no apparent differences in subjects with regard to radiotracer metabolism.

Analysis of PET brain data suggests:

- 1) Good initial brain penetrance and uptake with relatively fast washout.
- 2) Two tissue compartment models well-characterize the data, including good fits over the first 90 minutes post injection.
- 3) Delayed appearance of skull uptake affects quantitation when later time points are used, raising V_t - due to spillage into adjacent neocortical regions.

In summary, human validation studies of [18]F PBR111 demonstrate favorable metabolism and brain kinetics for kinetic modeling of TSPO receptor density (V_t or BPnd). There were no safety issues identified in any of the eleven subjects.

Conclusion

This report describes our scientific progress with our back-up TSPO PET agent 18F-PBR111 acquired in parallel projects evaluating neurodegenerative disease and suggests that we have adequate signal:noise properties to proceed with the approved PTSD study. All DOD and FDA regulatory requirements have been completed and we are now actively recruiting.

References

1. Moran, L.B., D.C. Duke, and M.B. Graeber, *The microglial gene regulatory network activated by interferon-gamma*. Journal of Neuroimmunology, 2007. **183**(1-2): p. 1.
2. Moran, L.B., D.C. Duke, F.E. Turkheimer, R.B. Banati, and M.B. Graeber, *Towards a transcriptome definition of microglial cells*. Neurogenetics, 2004. **5**(2): p. 95-108.
3. Rock, R.B., S. Hu, A. Deshpande, S. Munir, B.J. May, C.A. Baker, P.K. Peterson, and V. Kapur, *Transcriptional response of human microglial cells to interferon-[gamma]*. Genes Immun, 2005. **6**(8): p. 712.
4. Thomas, D.M., D.M. Francescutti-Verbeem, and D.M. Kuhn, *Gene expression profile of activated microglia under conditions associated with dopamine neuronal damage*. FASEB J., 2005: p. 05-4873jfe.
5. Beurdeley-Thomas, A., L. Miccoli, S. Oudard, B. Dutrillaux, and M.F. Poupon, *The peripheral benzodiazepine receptors: a review*. J Neurooncol, 2000. **46**(1): p. 45-56.
6. Venneti, S., B.J. Lopresti, and C.A. Wiley, *The peripheral benzodiazepine receptor (Translocator protein 18kDa) in microglia: from pathology to imaging*. Prog Neurobiol, 2006. **80**(6): p. 308-22.
7. Gavish, M., I. Bachman, R. Shoukrun, Y. Katz, L. Veenman, G. Weisinger, and A. Weizman, *Enigma of the peripheral benzodiazepine receptor*. Pharmacol Rev, 1999. **51**(4): p. 629-50.
8. Maeda, J., M. Higuchi, M. Inaji, B. Ji, E. Haneda, T. Okauchi, M.R. Zhang, K. Suzuki, and T. Suhara, *Phase-dependent roles of reactive microglia and astrocytes in nervous system injury as delineated by imaging of peripheral benzodiazepine receptor*. Brain Res, 2007. **1157**: p. 100-11.
9. Debruyne, J.C., J. Versijpt, K.J. Van Laere, F. De Vos, J. Keppens, K. Strijckmans, E. Achten, G. Slegers, R.A. Dierckx, J. Korf, and J.L. De Reuck, *PET visualization of microglia in multiple sclerosis patients using [11C]PK11195*. Eur J Neurol, 2003. **10**(3): p. 257-64.
10. Debruyne, J.C., K.J. Van Laere, J. Versijpt, F. De Vos, J.K. Eng, K. Strijckmans, P. Santens, E. Achten, G. Slegers, J. Korf, R.A. Dierckx, and J.L. De Reuck, *Semiquantification of the peripheral-type benzodiazepine ligand [11C]PK11195 in normal human brain and application in multiple sclerosis patients*. Acta Neurol Belg, 2002. **102**(3): p. 127-35.
11. Versijpt, J., J.C. Debruyne, K.J. Van Laere, F. De Vos, J. Keppens, K. Strijckmans, E. Achten, G. Slegers, R.A. Dierckx, J. Korf, and J.L. De Reuck, *Microglial imaging with positron emission tomography and atrophy measurements with magnetic resonance imaging in multiple sclerosis: a correlative study*. Mult Scler, 2005. **11**(2): p. 127-34.
12. Hammoud, D.A., C.J. Endres, A.R. Chander, T.R. Guilarte, D.F. Wong, N.C. Sacktor, J.C. McArthur, and M.G. Pomper, *Imaging glial cell activation with [11C]-R-PK11195 in patients with AIDS*. J Neurovirol, 2005. **11**(4): p. 346-55.
13. Chen, M.K. and T.R. Guilarte, *Imaging the peripheral benzodiazepine receptor response in central nervous system demyelination and remyelination*. Toxicol Sci, 2006. **91**(2): p. 532-9.
14. Chen, M.K., K. Baidoo, T. Verina, and T.R. Guilarte, *Peripheral benzodiazepine receptor imaging in CNS demyelination: functional implications of anatomical and cellular localization*. Brain, 2004. **127**(Pt 6): p. 1379-92.
15. Venneti, S., B.J. Lopresti, G. Wang, S.J. Bissel, C.A. Mathis, C.C. Meltzer, F. Boada, S. Capuano, 3rd, G.J. Kress, D.K. Davis, J. Ruszkiewicz, I.J. Reynolds, M. Murphey-Corb, A.M. Trichel, S.R. Wisniewski, and C.A. Wiley, *PET imaging of brain macrophages using the peripheral benzodiazepine receptor in a macaque model of neuroAIDS*. J Clin Invest, 2004. **113**(7): p. 981-9.
16. Mankowski, J.L., S.E. Queen, P.J. Tarwater, R.J. Adams, and T.R. Guilarte, *Elevated peripheral benzodiazepine receptor expression in simian immunodeficiency virus encephalitis*. J Neurovirol, 2003. **9**(1): p. 94-100.
17. Remington, L.T., A.A. Babcock, S.P. Zehntner, and T. Owens, *Microglial recruitment, activation, and proliferation in response to primary demyelination*. Am J Pathol, 2007. **170**(5): p. 1713-24.

18. Mattner, F., A. Katsifis, M. Staykova, P. Ballantyne, and D.O. Willenborg, *Evaluation of a radiolabelled peripheral benzodiazepine receptor ligand in the central nervous system inflammation of experimental autoimmune encephalomyelitis: a possible probe for imaging multiple sclerosis*. Eur J Nucl Med Mol Imaging, 2005. **32**(5): p. 557-63.
19. Bartels, A.L. and K.L. Leenders, *Neuroinflammation in the pathophysiology of Parkinson's disease: Evidence from animal models to human in vivo studies with [(11)C]-PK11195 PET*. Mov Disord, 2007.
20. Cagnin, A., R. Myers, R.N. Gunn, A.D. Lawrence, T. Stevens, G.W. Kreutzberg, T. Jones, and R.B. Banati, *In vivo visualization of activated glia by [(11)C] (R)-PK11195-PET following herpes encephalitis reveals projected neuronal damage beyond the primary focal lesion*. Brain, 2001. **124**(Pt 10): p. 2014-27.
21. Diorio, D., S.A. Welner, R.F. Butterworth, M.J. Meaney, and B.E. Suranyi-Cadotte, *Peripheral benzodiazepine binding sites in Alzheimer's disease frontal and temporal cortex*. Neurobiol Aging, 1991. **12**(3): p. 255-8.
22. Kropholler, M.A., R. Boellaard, B.N. van Berckel, A. Schuitemaker, R.W. Kloet, M.J. Lubberink, C. Jonker, P. Scheltens, and A.A. Lammertsma, *Evaluation of reference regions for (R)-[(11)C]PK11195 studies in Alzheimer's disease and Mild Cognitive Impairment*. J Cereb Blood Flow Metab, 2007. **21**.
23. Versijpt, J.J., F. Dumont, K.J. Van Laere, D. Decoo, P. Santens, K. Audenaert, E. Achten, G. Slegers, R.A. Dierckx, and J. Korf, *Assessment of neuroinflammation and microglial activation in Alzheimer's disease with radiolabelled PK11195 and single photon emission computed tomography. A pilot study*. Eur Neurol, 2003. **50**(1): p. 39-47.
24. Sapolsky, R.M., *Glucocorticoids and hippocampal atrophy in neuropsychiatric disorders*. Arch Gen Psychiatry, 2000. **57**(10): p. 925-35.
25. Gould, E., B.S. McEwen, P. Tanapat, L.A.M. Galea, and E. Fuchs, *Neurogenesis in the Dentate Gyrus of the Adult Tree Shrew Is Regulated by Psychosocial Stress and NMDA Receptor Activation*. J. Neurosci., 1997. **17**(7): p. 2492-2498.
26. Liston, C., M.M. Miller, D.S. Goldwater, J.J. Radley, A.B. Rocher, P.R. Hof, J.H. Morrison, and B.S. McEwen, *Stress-Induced Alterations in Prefrontal Cortical Dendritic Morphology Predict Selective Impairments in Perceptual Attentional Set-Shifting*. J. Neurosci., 2006. **26**(30): p. 7870-7874.
27. McEwen, B.S., *Physiology and Neurobiology of Stress and Adaptation: Central Role of the Brain*. Physiol. Rev., 2007. **87**(3): p. 873-904.
28. Bremner, J.D., *Stress and brain atrophy*. CNS Neurol Disord Drug Targets, 2006. **5**(5): p. 503-12.
29. Duman, R.S. and L.M. Monteggia, *A Neurotrophic Model for Stress-Related Mood Disorders*. Biological Psychiatry, 2006. **59**(12): p. 1116.
30. Sapolsky, R.M., *Stress, Glucocorticoids, and Damage to the Nervous System: The Current State of Confusion*. Stress, 1996. **1**(1): p. 1-19.
31. Sapolsky, R.M., *Glucocorticoids, stress, and their adverse neurological effects: relevance to aging*. Exp Gerontol, 1999. **34**(6): p. 721-32.
32. Sapolsky, R.M., H. Uno, C.S. Rebert, and C.E. Finch, *Hippocampal damage associated with prolonged glucocorticoid exposure in primates*. J Neurosci, 1990. **10**(9): p. 2897-902.
33. McEwen, B.S. and R.M. Sapolsky, *Stress and cognitive function*. Curr Opin Neurobiol, 1995. **5**(2): p. 205-16.
34. McEwen, B.S., R.E. Brinton, and R.M. Sapolsky, *Glucocorticoid receptors and behavior: implications for the stress response*. Adv Exp Med Biol, 1988. **245**: p. 35-45.
35. McIntosh, L.J. and R.M. Sapolsky, *Glucocorticoids may enhance oxygen radical-mediated neurotoxicity*. Neurotoxicology, 1996. **17**(3-4): p. 873-82.
36. Lehmann, J., R. Weizman, C.R. Pryce, S. Leschiner, I. Allmann, J. Feldon, and M. Gavish, *Peripheral benzodiazepine receptors in cerebral cortex, but not in internal organs, are increased following inescapable stress and subsequent avoidance/escape shuttle-box testing*. Brain Res, 1999. **851**(1-2): p. 141-7.
37. Bitran, D., D. Carlson, S. Leschiner, and M. Gavish, *Ovarian steroids and stress produce changes in peripheral benzodiazepine receptor density*. European Journal of Pharmacology, 1998. **361**(2-3): p. 235.
38. Drugan, R.C., P.V. Holmes, D.M. Scher, S. Luczak, H. Oh, and R.J. Ferland, *Environmentally induced changes in peripheral benzodiazepine receptors are stressor and tissue specific*. Pharmacology Biochemistry and Behavior, 1995. **50**(4): p. 551.
39. Drugan, R.C., P.V. Holmes, and A.P. Stringer, *Sexual dimorphism of stress-induced changes in renal peripheral benzodiazepine receptors in rat*. Neuropharmacology, 1991. **30**(4): p. 413-6.
40. Holmes, P.V., A.P. Stringer, and R.C. Drugan, *Impact of psychological dynamics of stress on the peripheral benzodiazepine receptor*. Pharmacol Biochem Behav, 1992. **42**(3): p. 437-44.

41. Gavish, M., N. Laor, M. Bidder, D. Fisher, O. Fonia, U. Muller, A. Reiss, L. Wolmer, L. Karp, and R. Weizman, *Altered platelet peripheral-type benzodiazepine receptor in posttraumatic stress disorder*. Neuropsychopharmacology, 1996. **14**(3): p. 181-6.
42. Pitman, R.K., *Hippocampal diminution in PTSD: More (or less?) than meets the eye*. Hippocampus, 2001. **11**(2): p. 73-74.
43. Pitman, R.K., M.W. Gilbertson, T.V. Gurvits, F.S. May, N.B. Lasko, L.J. Metzger, M.E. Shenton, R. Yehuda, and S.P. Orr, *Clarifying the origin of biological abnormalities in PTSD through the study of identical twins discordant for combat exposure*. Ann N Y Acad Sci, 2006. **1071**: p. 242-54.
44. Gulyas, B., C. Halldin, A. Vas, R.B. Banati, E. Shchukin, S. Finnema, J. Tarkainen, K. Tihanyi, G. Szilagyi, and L. Farde, *[11C]vinpocetine: a prospective peripheral benzodiazepine receptor ligand for primate PET studies*. J Neurol Sci, 2005. **229-230**: p. 219-23.
45. Gurvits, T.V., L.J. Metzger, N.B. Lasko, P.A. Cannistraro, A.S. Tarhan, M.W. Gilbertson, S.P. Orr, A.M. Charbonneau, M.M. Wedig, and R.K. Pitman, *Subtle neurologic compromise as a vulnerability factor for combat-related posttraumatic stress disorder: results of a twin study*. Arch Gen Psychiatry, 2006. **63**(5): p. 571-6.
46. Sapolsky, R.M., *Chickens, eggs and hippocampal atrophy*. Nat Neurosci, 2002. **5**(11): p. 1111-3.
47. Bremner, J.D., M. Vythilingam, E. Vermetten, S.M. Southwick, T. McGlashan, A. Nazeer, S. Khan, L.V. Vaccarino, R. Soufer, P.K. Garg, C.K. Ng, L.H. Staib, J.S. Duncan, and D.S. Charney, *MRI and PET study of deficits in hippocampal structure and function in women with childhood sexual abuse and posttraumatic stress disorder*. Am J Psychiatry, 2003. **160**(5): p. 924-32.
48. Sapolsky, R.M., *Atrophy of the hippocampus in posttraumatic stress disorder: how and when?* Hippocampus, 2001. **11**(2): p. 90-1.
49. Bremner, J.D., P. Randall, T.M. Scott, R.A. Bronen, J.P. Seibyl, S.M. Southwick, R.C. Delaney, G. McCarthy, D.S. Charney, and R.B. Innis, *MRI-based measurement of hippocampal volume in patients with combat-related posttraumatic stress disorder*. Am J Psychiatry, 1995. **152**(7): p. 973-81.
50. Bremner, J.D., R.B. Innis, S.M. Southwick, L. Staib, S. Zoghbi, and D.S. Charney, *Decreased benzodiazepine receptor binding in prefrontal cortex in combat-related posttraumatic stress disorder*. Am J Psychiatry, 2000. **157**(7): p. 1120-6.
51. Fujita, M., S.M. Southwick, C.C. Denucci, S.S. Zoghbi, M.S. Dillon, R.M. Baldwin, A. Bozkurt, A. Kugaya, N.P. Verhoeff, J.P. Seibyl, and R.B. Innis, *Central type benzodiazepine receptors in Gulf War veterans with posttraumatic stress disorder*. Biol Psychiatry, 2004. **56**(2): p. 95-100.
52. Seal, K.H., D. Bertenthal, C.R. Miner, S. Sen, and C. Marmar, *Bringing the war back home: mental health disorders among 103,788 US veterans returning from Iraq and Afghanistan seen at Department of Veterans Affairs facilities*. Arch Intern Med, 2007. **167**(5): p. 476-82.
53. Hoge, C.W., A. Terhakopian, C.A. Castro, S.C. Messer, and C.C. Engel, *Association of posttraumatic stress disorder with somatic symptoms, health care visits, and absenteeism among Iraq war veterans*. Am J Psychiatry, 2007. **164**(1): p. 150-3.
54. Friedman, M.J., *Acknowledging the psychiatric cost of war*. N Engl J Med, 2004. **351**(1): p. 75-7.
55. Kudler, H., *Chronic stress and adaptation*. Am J Psychiatry, 2006. **163**(3): p. 552-3; author reply 553.
56. Rona, R.J., R. Hooper, M. Jones, L. Hull, T. Browne, O. Horn, D. Murphy, M. Hotopf, and S. Wessely, *Mental health screening in armed forces before the Iraq war and prevention of subsequent psychological morbidity: follow-up study*. Bmj, 2006. **333**(7576): p. 991.
57. Rona, R.J., K.C. Hyams, and S. Wessely, *Screening for psychological illness in military personnel*. Jama, 2005. **293**(10): p. 1257-60.
58. Hoge, C.W., C.A. Castro, S.C. Messer, D. McGurk, D.I. Cotting, and R.L. Koffman, *Combat duty in Iraq and Afghanistan, mental health problems, and barriers to care*. N Engl J Med, 2004. **351**(1): p. 13-22.
59. Turkheimer, F.E., P. Edison, N. Pavese, F. Roncaroli, A.N. Anderson, A. Hammers, A. Gerhard, R. Hinz, Y.F. Tai, and D.J. Brooks, *Reference and target region modeling of [11C]-(R)-PK11195 brain studies*. J Nucl Med, 2007. **48**(1): p. 158-67.
60. Schuitmaker, A., B.N. van Berckel, M.A. Kroppoller, D.J. Veltman, P. Scheltens, C. Jonker, A.A. Lammertsma, and R. Boellaard, *SPM analysis of parametric (R)-[11C]PK11195 binding images: plasma input versus reference tissue parametric methods*. Neuroimage, 2007. **35**(4): p. 1473-9.
61. Imaizumi, M., H.J. Kim, S.S. Zoghbi, E. Briard, J. Hong, J.L. Musachio, C. Ruetzler, D.M. Chuang, V.W. Pike, R.B. Innis, and M. Fujita, *PET imaging with [11C]PBR28 can localize and quantify upregulated peripheral benzodiazepine receptors associated with cerebral ischemia in rat*. Neurosci Lett, 2007. **411**(3): p. 200-5.

62. Shinotoh, H., *Neuroimaging of PD, PSP, CBD and MSA-PET and SPECT studies*. J Neurol, 2006. **253 Suppl 3**: p. iii30-iii34.
63. Dumont, F., F. De Vos, J. Versijpt, H.M. Jansen, J. Korf, R.A. Dierckx, and G. Slegers, *In vivo evaluation in mice and metabolism in blood of human volunteers of [123I]iodo-PK11195: a possible single-photon emission tomography tracer for visualization of inflammation*. Eur J Nucl Med, 1999. **26**(3): p. 194-200.
64. Gildersleeve, D.L., M.E. Van Dort, J.W. Johnson, P.S. Sherman, and D.M. Wieland, *Synthesis and evaluation of [123I]-iodo-PK11195 for mapping peripheral-type benzodiazepine receptors (omega 3) in heart*. Nucl Med Biol, 1996. **23**(1): p. 23-8.
65. Boutin, H., F. Chauveau, C. Thominaux, M.C. Gregoire, M.L. James, R. Trebossen, P. Hantraye, F. Dolle, B. Tavitian, and M. Kassiou, *11C-DPA-713: a novel peripheral benzodiazepine receptor PET ligand for in vivo imaging of neuroinflammation*. J Nucl Med, 2007. **48**(4): p. 573-81.
66. Seibyl, J.P., E. Wallace, E.O. Smith, M. Stabin, R.M. Baldwin, S. Zoghbi, Y. Zea-Ponce, Y. Gao, W.Y. Zhang, J.L. Neumeyer, and et al., *Whole-body biodistribution, radiation absorbed dose and brain SPECT imaging with iodine-123- beta-CIT in healthy human subjects*. J Nucl Med, 1994. **35**(5): p. 764-70.
67. Dey, H.M., J.P. Seibyl, J.B. Stubbs, S.S. Zoghbi, R.M. Baldwin, E.O. Smith, I.G. Zubal, Y. Zea-Ponce, C. Olson, D.S. Charney, and et al., *Human biodistribution and dosimetry of the SPECT benzodiazepine receptor radioligand iodine-123- iomazenil*. J Nucl Med, 1994. **35**(3): p. 399-404.
68. Fujita, M., J.P. Seibyl, D.B. Vaupel, G. Tamagnan, M. Early, S.S. Zoghbi, R.M. Baldwin, A.G. Horti, N.A. Kore, A.G. Mukhin, S. Khan, A. Bozkurt, A.S. Kimes, E.D. London, and R.B. Innis, *Whole-body biodistribution, radiation absorbed dose, and brain SPET imaging with [123I]5-i-A-85380 in healthy human subjects*. Eur J Nucl Med Mol Imaging, 2002. **29**(2): p. 183-90.
69. Abi-Dargham, A., R.B. Innis, G. Wisniewski, R.M. Baldwin, J.L. Neumeyer, and J.P. Seibyl, *Human biodistribution and dosimetry of iodine-123-fluoroalkyl analogs of beta-CIT*. Eur J Nucl Med, 1997. **24**(11): p. 1422-5. 23
70. van Dyck, C.H., J.P. Seibyl, J.B. Stubbs, S. Zoghbi, G. Wisniewski, R.M. Baldwin, Y. Zea-Ponce, D.S. Charney, P.B. Hoffer, and R.B. Innis, *Human biodistribution and dosimetry of the SPECT D2 dopamine receptor radioligand [123I]IBF*. Nucl Med Biol, 1996. **23**(1): p. 9-16.

Appendix

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